

Tests on the Reliability and Throughput Performance of hSMD

Computed Values of Binding Free Energy *vs* Experimental Data

Liao Y Chen, PhD

Department of Physics, UTSA

| Binding complex (PDB code of complex/protein) | Model systems (# of atoms) | Computing time: hours on # of CPU- cores* | hSMD prediction in kcal/mol | Experimentally measured value in kcal/mol (method) | Other computed results in kcal/mol (method) |
|---|-------------------------------------|--|--------------------------------------|--|---|
| 1. OCA-LGB (3NQ9) | 96K | 62 on 480 | -5.76 ¹ | -5.57 (FS) ² | -2.66 (GBSA) ³ |
| 2. DKA-LGB (3NQ3) | 96K | 64 on 480 | -4.44 ⁴ | -5.22 (FS) ² | -4.44 (GBSA) ³ |
| 3. DAO-LGB (3UEU) | 96K | 66 on 480 | -6.89 ⁴ | -7.24 (ITC) ⁵ | -12.37 (GBSA) ³ |
| 4. MYR-LGB (3UEV) | 96K | 68 on 480 | -8.57 ⁴ | -8.14 (ITC) ⁵ | -12.66 (GBSA) ³ |
| 5. PLM-LGB (3UEW) | 96K | 70 on 480 | -8.99 ⁴ | -8.72 (ITC) ⁵ | -20.92 (GBSA) ³ |
| 6. GSH-SjGST (1U87) | 115K | 88 on 512 | -7.0 ¹ | -6.75 (ITC) ⁶ | - |
| 7. 8ST-VEGFR1 (3HNG) | 95K | 63 on 256 | -9.2 ⁷ | -8.3 (IC50) ⁸ | - |
| 8. Tuftsin-NRP-1 (2ORZ) | 125K | 53 on 512 | -7.2 | -7.2 (ITC) ⁹ | -32 (JE-SMD) |
| 9. Ras-RalGDS (1LFD) | 96K | 72 on 400 | -9.2 ¹⁰ | -8.4 (ITC) ¹¹ | -19.5 (GBSA) ¹² |
| 10. Barnase- Barstar (1BRS) | 97K | 72 on 320 | -16.8 ¹⁰ | -16.0 (ITC) ¹³ | -21.0 (wt, GRC) ¹⁴ |
| 11. GOL-GLPF (1FX8) | 78K | 58 on 320 | -3.5 ¹⁵ | <-2.8 (XRD) ¹⁶ <-2.7 (IC50) ¹⁷ | - |
| 12. GOL-PfAQP (3C02) | 150K | 58 on 640 | -3.8 ¹⁸ | <-2.8 (XRD) ¹⁹ <<-1.1 (IC50) ^{18, 20} | - |
| 13. MRYPfAQP (3C02) | 150K | 58 on 640 | -8.4 ¹⁸ | TBV ^{&} | - |
| 14. AZM-AQP4 (3GD8) | 73K | 58 on 320 | -3.7 ²¹ | -3.4 (IC50) ²² | -6.5 (Virtual Screening) ²³ -3.2 (GRC) ²⁴ |
| 15. EDO-AQP4 (3GD8) | 73K | 88 on 256 | -1.5 ²¹ | TBV | - |
| 16. PDO-AQP4 (3GD8) | 73K | 88 on 256 | -4.75 ²¹ | TBV | - |

| | | | | | |
|----------------------|------|------------|---------------------|--|---|
| 17. AZM-caII (3HS4) | 61K | 71 on 200 | -11.1 ²⁵ | -10.8 (multiple, e.g. IC50 ²⁶) | - |
| 18. J0Z-T4L (3HUK) | 97K | 120 on 480 | -4.6 ²⁷ | -4.5 (ITC) ²⁸ | -1.3 (apo) -3.9 (holo) (FEP) ²⁸ |
| 19. 265-T4L (2RBO) | 97K | 120 on 480 | -4.9 ²⁷ | -4.9 (ITC) ²⁸ | -5.7 (apo) -11.4 (holo) (FEP) ²⁸ |
| 20. BNZ-T4L (4I7) | 97K | 120 on 480 | -4.8 ²⁷ | -4.5 (ITC) ²⁹ | - |
| 21. RTL-CRBP1 (5HBS) | 103K | 132 on 480 | -12.6 ²⁷ | 10.9 (ITC) ³⁰ | - |

*Complexes 1-5 were computed on TACC supercomputers Lonestar, Complexes 6-8, 15, and 16 on Stampede, and Complexes 9-14 and 17 on Maverick all **without invoking GPU or MIC acceleration**. For comparison, Complex 17 was also computed on a single node of Maverick (20 CPU-cores) **with GPU** acceleration took 198 hours, a longer time but at a lower computing cost of about 4K SUs which is in contrast 14 K SUs on 200 CPU-cores without GPU.

&TBV: To be validated *in vitro/vivo*.

References

- Chen, L. Y. Hybrid Steered Molecular Dynamics Approach to Computing Absolute Binding Free Energy of Ligand–Protein Complexes: A Brute Force Approach That Is Fast and Accurate. *J Chem Theory Comput* **2015**, 11, 1928-1938.
- Loch, J.; Polit, A.; Górecki, A.; Bonarek, P.; Kurpiewska, K.; Dziedzicka-Wasylewska, M.; Lewiński, K. Two modes of fatty acid binding to bovine β -lactoglobulin—crystallographic and spectroscopic studies. *J. Mol. Recognit.* **2011**, 24, 341-349.
- Bello, M. Binding free energy calculations between bovine β -lactoglobulin and four fatty acids using the MMGBSA Method. *Biopolymers* **2014**, 101, 1010-1018.
- Yi, C.; Wambo, T. O. Factors affecting the interactions between beta-lactoglobulin and fatty acids as revealed in molecular dynamics simulations. *Physical Chemistry Chemical Physics* **2015**.
- Loch, J. I.; Polit, A.; Bonarek, P.; Olszewska, D.; Kurpiewska, K.; Dziedzicka-Wasylewska, M.; Lewiński, K. Structural and thermodynamic studies of binding saturated fatty acids to bovine β -lactoglobulin. *Int. J. Biol. Macromol.* **2012**, 50, 1095-1102.
- Andújar-Sánchez, M.; Clemente-Jiménez, J. M.; amp; x; Las Heras-Vázquez, F. J.; Rodri; amp; x; guez-Vico, F.; Cámara-Artigas, A.; Jara-Pérez, V. Thermodynamics of glutathione binding to the tyrosine 7 to phenylalanine mutant of glutathione S-transferase from *Schistosoma japonicum*. *Int. J. Biol. Macromol.* **2003**, 32, 77-82.
- Chen, L. Y. Computing membrane-AQP5-phosphatidylserine binding affinities with hybrid steered molecular dynamics approach. *Mol. Membr. Biol.* **2015**.
- Furet, P.; Bold, G.; Hofmann, F.; Manley, P.; Meyer, T.; Altmann, K.-H. Identification of a new chemical class of potent angiogenesis inhibitors based on conformational considerations and database searching. *Bioorg. Med. Chem. Lett.* **2003**, 13, 2967-2971.
- von Wronski, M. A.; Raju, N.; Pillai, R.; Bogdan, N. J.; Marinelli, E. R.; Nanjappan, P.; Ramalingam, K.; Arunachalam, T.; Eaton, S.; Linder, K. E.; Yan, F.; Pochon, S.; Tweedle, M. F.; Nunn, A. D. Tuftsin Binds

Neuropilin-1 through a Sequence Similar to That Encoded by Exon 8 of Vascular Endothelial Growth Factor. *J. Biol. Chem.* **2006**, 281, 5702-5710.

10. Rodriguez, R. A.; Yu, L.; Chen, L. Y. Computing Protein-Protein Association Affinity with Hybrid Steered Molecular Dynamics. *J. Chem. Theory Comput.* **2015**.
11. Rudolph, M. G.; Linnemann, T.; Grünewald, P.; Wittinghofer, A.; Vetter, I. R.; Herrmann, C. Thermodynamics of Ras/Effector and Cdc42/Effector Interactions Probed by Isothermal Titration Calorimetry. *J. Biol. Chem.* **2001**, 276, 23914-23921.
12. Gohlke, H.; Kiel, C.; Case, D. A. Insights into Protein-Protein Binding by Binding Free Energy Calculation and Free Energy Decomposition for the Ras-Raf and Ras-RalGDS Complexes. *J. Mol. Biol.* **2003**, 330, 891-913.
13. Buckle, A. M.; Schreiber, G.; Fersht, A. R. Protein-protein recognition: Crystal structural analysis of a barnase-barstar complex at 2.0-Å resolution. *Biochemistry* **1994**, 33, 8878-8889.
14. Gumbart, J. C.; Roux, B.; Chipot, C. Efficient Determination of Protein-Protein Standard Binding Free Energies from First Principles. *J. Chem. Theory Comput.* **2013**, 9, 3789-3798.
15. Chen, L. Y. Glycerol modulates water permeation through Escherichia coli aquaglyceroporin GlpF. *Biochimica Et Biophysica Acta-Biomembranes* **2013**, 1828, 1786-1793.
16. Fu, D.; Libson, A.; Miercke, L. J. W.; Weitzman, C.; Nollert, P.; Krucinski, J.; Stroud, R. M. Structure of a Glycerol-Conducting Channel and the Basis for Its Selectivity. *Science* **2000**, 290, 481-486.
17. Borgnia, M. J.; Agre, P. Reconstitution and functional comparison of purified GlpF and AqpZ, the glycerol and water channels from Escherichia coli. *Proceedings of the National Academy of Sciences* **2001**, 98, 2888-2893.
18. Chen, L. Y. Erythritol predicted to inhibit permeation of water and solutes through the conducting pore of P. falciparum aquaporin. *Biophysical Chemistry* **2015**, 198, 14-21.
19. Hansen, M.; Kun, J. F. J.; Schultz, J. E.; Beitz, E. A Single, Bi-functional Aquaglyceroporin in Blood-stage Plasmodium falciparum Malaria Parasites. *J. Biol. Chem.* **2002**, 277, 4874-4882.
20. Song, J.; Almasalmeh, A.; Krenc, D.; Beitz, E. Molar concentrations of sorbitol and polyethylene glycol inhibit the Plasmodium aquaglyceroporin but not that of E. coli: involvement of the channel vestibules. *Biochim. Biophys. Acta* **2012**, 1818, 1218-24.
21. Yu, L.; Villarreal, O. D.; Laurie Chen, L.; Chen, L. Y. 1,3-Propanediol binds inside the water-conducting pore of aquaporin 4: Does this efficacious inhibitor have sufficient potency? *J Syst Integr Neurosci* **2016**, 2, 91-98.
22. Ho, J. D.; Yeh, R.; Sandstrom, A.; Chorny, I.; Harries, W. E. C.; Robbins, R. A.; Miercke, L. J. W.; Stroud, R. M. Crystal structure of human aquaporin 4 at 1.8 Å and its mechanism of conductance. *Proceedings of the National Academy of Sciences* **2009**, 106, 7437-7442.
23. Huber, V. J.; Tsujita, M.; Kwee, I. L.; Nakada, T. Inhibition of Aquaporin 4 by antiepileptic drugs. *Biorg. Med. Chem.* **2009**, 17, 418-424.
24. Villarreal, O. D.; Rodriguez, R. A.; Yu, L.; Wambo, T. O. Molecular dynamics simulations on the effect of size and shape on the interactions between negative Au18(SR)14, Au102(SR)44 and Au144(SR)60 nanoparticles in physiological saline. *Colloids and Surfaces A: Physicochemical and Engineering Aspects* **2016**, 503, 70-78.
25. Wambo, T. O.; Chen, L. Y.; McHardy, S. F.; Tsin, A. T. Molecular dynamics study of human carbonic anhydrase II in complex with Zn²⁺ and acetazolamide on the basis of all-atom force field simulations. *Biophys. Chem.* **2016**, 214-215, 54-60.
26. Wilkinson, B. L.; Bornaghi, L. F.; Houston, T. A.; Innocenti, A.; Vullo, D.; Supuran, C. T.; Poulsen, S.-A. Carbonic Anhydrase Inhibitors: Inhibition of Isozymes I, II, and IX with Triazole-Linked O-Glycosides of Benzene Sulfonamides. *J. Med. Chem.* **2007**, 50, 1651-1657.

27. Villarreal, O. D.; Yu, L.; Rodriguez, R. A.; Chen, L. Y. Computing the binding affinity of a ligand buried deep inside a protein with the hybrid steered molecular dynamics. *Biochem. Biophys. Res. Commun.* **2017**, 483, 203-208.
28. Boyce, S. E.; Mobley, D. L.; Rocklin, G. J.; Graves, A. P.; Dill, K. A.; Shoichet, B. K. Predicting Ligand Binding Affinity with Alchemical Free Energy Methods in a Polar Model Binding Site. *J. Mol. Biol.* **2009**, 394, 747-763.
29. Merski, M.; Shoichet, B. K. The Impact of Introducing a Histidine into an Apolar Cavity Site on Docking and Ligand Recognition. *J. Med. Chem.* **2013**, 56, 2874-2884.
30. Silvaroli, J. A.; Arne, J. M.; Chelstowska, S.; Kiser, P. D.; Banerjee, S.; Golczak, M. Ligand Binding Induces Conformational Changes in Human Cellular Retinol-binding Protein 1 (CRBP1) Revealed by Atomic Resolution Crystal Structures. *J. Biol. Chem.* **2016**, 291, 8528-8540.